Supplementary materials

Supplementary	Table	1. 16S	and 23S rRNA gene-target	ed primers u	used in this s	tudy
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Target bacteria*	Primer	Sequence (5' - 3')	Ref
Clostridium coccoides group	g-Ccoc-F	AAATGACGGTACCTGACTAA	5
	g-Ccoc-R	CTTTGAGTTTCATTCTTGCGAA	
Clostridium leptum subgroup	sg-Clept-F	GCACAAGCAGTGGAGT	6
	sg-Clept-R3	CTTCCTCCGTTTTGTCAA	
Bacteroides fragilis group	g-Bfra-F2	AYAGCCTTTCGAAAGRAAGAT	7
	g-Bfra-R	CCAGTATCAACTGCAATTTTA	5
Bifidobacterium	g-Bifid-F	CTCCTGGAAACGGGTGG	5
	g-Bifid-R	GGTGTTCTTCCCGATATCTACA	
Atopobium cluster	g-Atopo-F	GGGTTGAGAGACCGACC	6
	g-Atopo-R	CGGRGCTTCTTCTGCAGG	
Prevotella	g-Prevo-F	CACRGTAAACGATGGATGCC	5
	g-Prevo-R	GGTCGGGTTGCAGACC	
Clostridium perfringens	s-Clper-F	GGGGGTTTCAACACCTCC	2
1 7 0	CIPER-R	GCAAGGGATGTCAAGTGT	4
Lactobacillus gasseri subgroup	sg-Lgas-F	GATGCATAGCCGAGTTGAGAGACTGAT	2
0 0 1	sg-Lgas-R	TAAAGGCCAGTTACTACCTCTATCC	
Lactobacillus brevis	s-Lbre-F	ATTTTGTTTGAAAGGTGGCTTCGG	2
	s-Lbre-R	ACCCTTGAACAGTTACTCTCAAAGG	_
Lactobacillus casei subgroup	sg-L cas-F	ACCGCATGGTTCTTGGC	2
r	sg-Lcas-R	CCGACAACAGTTACTCTGCC	_
Lactobacillus fermentum	LFer-1	CCTGATTGATTTTGGTCGCCAAC	2
Lacrooucinas jei mennam	LFer-2	ACGTATGAACAGTTACTCTCATACGT	-
Lactobacillus fructivorans	s-Lfru-F	TGCGCCTAATGATAGTTGA	2
	s-Lfru-R	GATACCGTCGCGACGTGAG	-
Lactobacillus plantarum subgroup	sg-Lpla-F	CTCTGGTATTGATTGGTGCTTGCAT	2
Baelobaellins plantarium subgroup	sg-Lpla P	GTTCGCCACTCACTCAAATGTAAA	-
Lactobacillus reuteri subgroup	sg-Lpiu R	GAACGCAYTGGCCCAA	2
Euclobucilius remeri subgroup	sg-Lreu-P	TCCATTGTGGCCGATCAGT	2
Lactobacillus ruminis subgroup	sg-Lieu-K	CACCGAATGCTTGCAYTCACC	2
Euclobacilius ruminis subgroup	sg-Lrum-P	GCCGCGGGTCCATCCAAAA	2
Lactobacillus sakei subgroup	sg-Lium-K	CATAAAACCTAMCACCGCATGG	2
Laciobacilius sakei subgroup	sg-Lsak-P	TCAGTTACTATCAGATACPTTCTTCTC	2
Enterobacteriaceae	5g-LSak-K En-lsu-3E		1
Enterobacternaceae	En-lsu-3'P	TCAAGGACCAGTGTTCAGTGTC	1
Fritaraaaaus	g Engog E		С
Emerococcus	g-Elicoc-F		2
Stanhulogoggus	g-Encoc-K		С
Siaphylococcus	g-Staph-F		2
Stranto accourt	g-Stapti-K		2
Sirepiococcus	g-Su-F		3
Providementa	g-Sit-K DSD7E		1
r seudomonas			1

*Specific primer sets were developed using 16S rDNA sequences, except for En-lsu-3F/3'R, and g-Str-F/R, which targeted 23S rDNA.

References

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- 4. Kikuchi E, Miyamoto Y, Narushima S, et al. Design of species specific primers to identify 13 species of *Clostridium* harbored in human intestinal tracts. *Microbiol Immunol* 2002; 46: 353–358.
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	db/+		db/db		polymyxin B-treated <i>db/db</i>	
	Bacterial count	Detection	Bacterial count	Detection	Bacterial count	Detection
	(log ₁₀ cells/g)	rate (%)	(log ₁₀ cells/g)	rate (%)	(log ₁₀ cells/g)	rate (%)
Total bacteria	9.3 ± 0.3		9.5 ± 0.3		9.5 ± 0.1	
Obligate anaerobes						
Clostridium coccoides group	8.3 ± 0.4	100	8.8 ± 0.4	80	8.5 ± 0.2	100
Clostridium leptum subgroup	8.5 ± 0.4	100	8.8 ± 0.2	80	8.7 ± 0.2	100
Bacteroides fragilis group	8.4 ± 0.4	100	8.6 ± 0.1	100	8.7 ± 0.3	100
Bifidobacterium	8.2 ± 1.0	100	<5.0	0	8.6 ± 0.6	40
Atopobium cluster	5.5 ± 0.7	60	6.6 ± 0.6	100	7.1 ± 0.5	100
Prevotella	<5.0	0	<5.0	0	<5.0	0
Clostridium perfringens	5.2 ± 0.1	60	4.4	20	4.4 ± 0	40
Facultative anaerobes						
Total Lactobacillus	8.9 ± 0.3		9.0 ± 0.3		9.2 ± 0.1	
Lactobacillus gasseri subgroup	8.5 ± 0.1	80	8.5 ± 0.4	40	8.9 ± 0.1	60
Lactobacillus brevis	2.7 ± 0.2	100	3.5 ± 0.2	100	3.3 ± 0.2	100
Lactobacillus casei subgroup	<3.0	0	<3.0	0	3.1	20
Lactobacillus fermentum	<4.0	0	<4.0	0	<4.0	0
Lactobacillus fructivorans	<2.4	0	<2.4	0	<2.4	0
Lactobacillus plantarum subgroup	3.5 ± 0.2	100	3.7 ± 0.3	100	3.5 ± 0.1	100
Lactobacillus reuteri subgroup	8.6 ± 0.4	100	8.8 ± 0.4	100	9.0 ± 0.2	100
Lactobacillus ruminis subgroup	7.0 ± 0.4	100	7.8 ± 0.3	100	7.9 ± 0.6	100
Lactobacillus sakei subgroup	3.7	20	2.4	20	<2.3	0
Enterobacteriaceae	5.1 ± 0.3	60	7.5 ± 0.6	100	5.5 ± 0.7	100
Enterococcus	6.6 ± 0.3	100	8.7 ± 0.5	100	7.5 ± 0.5	100
Staphylococcus	5.7 ± 0.4	100	5.7 ± 0.2	100	5.6 ± 0.3	100
Streptococcus	7.1 ± 0.7	100	6.3 ± 0.5	80	7.4 ± 0.2	100
Aerobes						
Pseudomonas	<3.0	0	<3.0	0	<3.0	0

Supplementary Table 2. Fecal bacterial counts and detection rates before middle cerebral artery occlusion in db/+, db/db, and polymyxin B-treated db/db mice (n = 5 per group).

Data are presented as mean \pm SD.

	Before MCAO	After MCAO	
	LPS (E	U/mL)	P value
db/+	2.1 ± 1.1	6.7 ± 2.3	<0.01
db/db	6.5 ± 0.9	19.6 ± 0.9	< 0.001
polymyxin B-treated <i>db/db</i>	3.8 ± 1.1	10.1 ± 2.9	< 0.001

Supplementary Table 3. Comparisons of plasma lipopolysaccharide (LPS) levels before and after middle cerebral artery occlusion (MCAO) in db/+, db/db, and polymyxin B-treated db/db mice (n = 6-9 per group).

Data are presented as mean \pm SD.

luorescein isothiocyanate (FITC)-dextran translocation before and after middle cerebral artery occlusion							
(MCAO) in $db/+$, db	<i>b/db</i> , and polymyxin B-treated <i>d</i>	b/db mice (n = 5 per group).					
	Before MCAO	After MCAO					
	FITC-dextran i	FITC-dextran in blood (ng/mL)					
db/+	12.2 ± 9.2	27.4 ± 8.2	< 0.05				

 108.8 ± 7.1

 46.5 ± 4.3

< 0.001

< 0.01

 32.7 ± 7.1

 17.6 ± 4.3

Supplementary Table 4. Comparisons of intestinal permeability assessed by quantitative analysis of

Data are presented as mean \pm SD.

polymyxin B-treated db/db

db/db

	db/+		db/db		polymyxin B-treated <i>db/db</i>	
	Bacterial count	Detection	Bacterial count	Detection	Bacterial count	Detection
	(log ₁₀ cells/g)	rate (%)	(log ₁₀ cells/g)	rate (%)	(log ₁₀ cells/g)	rate (%)
Total bacteria	9.8 ± 0.1		9.7 ± 0.1		10.0 ± 0.2	
Obligate anaerobes						
Clostridium coccoides group	9.0 ± 0.1	100	8.9 ± 0.2	100	9.1 ± 0.3	100
Clostridium leptum subgroup	8.9 ± 0.2	100	8.9 ± 0.3	100	8.9 ± 0.4	100
Bacteroides fragilis group	8.6 ± 0.2	100	8.9 ± 0.2	100	8.7 ± 0.3	100
Bifidobacterium	<4.9	0	<4.9	0	<4.9	0
Atopobium cluster	<5.0	0	<5.0	0	<5.0	0
Prevotella	<5.0	0	<5.0	0	<5.0	0
Clostridium perfringens	<2.3	0	<2.3	0	<2.3	0
Facultative anaerobes						
Total Lactobacillus	9.7 ± 0.2		9.3 ± 0.1		9.8 ± 0.1	
Lactobacillus gasseri subgroup	9.3 ± 0.3	100	8.5 ± 0.6	100	9.4 ± 0.1	100
Lactobacillus brevis	3.6 ± 0.2	100	4.2 ± 0.3	100	4.0 ± 0.2	100
Lactobacillus casei subgroup	2.9	20	3.2 ± 0.2	80	2.9	20
Lactobacillus fermentum	<4.0	0	4.4	20	<4.0	0
Lactobacillus fructivorans	2.4 ± 0.1	40	<2.3	0	<2.3	0
Lactobacillus plantarum subgroup	3.5 ± 0.5	100	4.0 ± 0.3	100	3.9 ± 0.2	100
Lactobacillus reuteri subgroup	9.0 ± 0.2	100	8.6 ± 0.3	100	9.2 ± 0.1	100
Lactobacillus ruminis subgroup	9.1 ± 0.2	100	9.0 ± 0.2	100	9.3 ± 0.2	100
Lactobacillus sakei subgroup	2.6	20	2.8 ± 0.5	60	2.6 ± 0	40
Enterobacteriaceae	<4.0	0	5.1	20	6.1	20
Enterococcus	7.9 ± 0.2	100	7.9 ± 0.4	100	8.5 ± 0.3	100
Staphylococcus	5.6 ± 0.3	100	5.7 ± 0.4	100	5.0 ± 0.3	100
Streptococcus	7.4 ± 0.2	100	7.9 ± 0.4	100	7.8 ± 0.5	100
Aerobes						
Pseudomonas	<2.9	0	<2.9	0	<2.9	0

Supplementary Table 5. Fecal bacterial counts and detection rates after middle cerebral artery occlusion in db/+, db/db, and polymyxin B-treated db/db mice (n = 5 per group).

Data are presented as mean \pm SD.

Supplementary Figure 1



Supplementary Figure 1. Fecal *Lactobacillus* counts (a) before middle cerebral artery occlusion (MCAO) and (b) 24 h after MCAO analyzed by rRNA-targeted quantitative reverse transcription PCR in db/+, db/db, and polymyxin B (PL-B)-treated db/db mice (n = 5 per group). Bacterial counts below the threshold of detection were not plotted. Data are shown as mean \pm SD.



Supplementary Figure 2. Relative abundance of predominant bacteria before middle cerebral artery occlusion in db/+, db/db, and polymyxin B (PL-B)-treated db/db mice (n = 5 per group).





Supplementary Figure 3. Physiological parameters. (a) Weight loss (baseline – 24 h after middle cerebral artery occlusion (MCAO)) (n = 5 per group) and (b) temporal changes in regional cerebral blood flow (rCBF) in ipsilateral hemispheres (n = 5 per group) in db/+, db/db, and polymyxin B (PL-B)-treated db/db mice. Data are shown as mean \pm SD.

Supplementary Figure 4



Supplementary Figure 4. Comparisons of fecal bacterial counts before and 24 h after middle cerebral

artery occlusion (MCAO) analyzed by rRNA-targeted quantitative reverse transcription PCR in db/+, db/db, and polymyxin B (PL-B)-treated db/db mice (n = 5 per group). Bacterial counts below the threshold of detection were not plotted. Data are shown as mean \pm SD. *P < 0.05, **P < 0.01, and ***P < 0.001.





Supplementary Figure 5. Effects of polymyxin B (PL-B) on stroke outcome in db/+ mice. (a) Plasma lipopolysaccharide (LPS) levels 24 h after middle cerebral artery occlusion (MCAO) (n = 5 per group). (b) Intestinal permeability 3 h after MCAO (n = 5 per group). (c) Quantification of infarct volumes 24 h after

MCAO in db/+ and polymyxin B-treated db/+ mice. Total hemispheric infarct volumes were corrected for edema (n = 5 per group). Data are shown as mean \pm SD. (d) Modified neurological severity scores (NSS) at 1 h and 24 h after MCAO in db/+ and polymyxin B-treated db/+ mice (n = 5 per group). Data are shown as median and interquartile range. (e) Fecal bacterial counts 24 h after MCAO analyzed by rRNA-targeted quantitative reverse transcription PCR in db/+ and polymyxin B-treated db/+ mice (n = 5 per group). Bacterial counts below the threshold of detection were not plotted. Data are shown as mean \pm SD.



Supplementary Figure 6. Expression of lipopolysaccharide (LPS), toll-like receptor 4 (TLR4), and inflammatory cytokines in non-ischemic hemispheres 24 h after middle cerebral artery occlusion. Western blot analyses of (a) LPS and (b) TLR4 expression. (c) Enzyme-linked immunoassay results for tumor necrosis factor (TNF)- α , interleukin (IL)-1 β , and IL-6. All data were analyzed in *db/+*, *db/db*, and polymyxin B (PL-B)-treated *db/db* mice sacrificed 24 h after middle cerebral artery occlusion (n = 4–5 per group). Data are shown as mean ± SD.